# Non linear genotype-phenotype mapping: RNA landscapes

Course computational biology 2025; Paulien Hogeweg; Theoretical Biology and Bioinformatics Grp Utrecht University Classical Population Dynamic replicator models + Space or vesicles ( emergent or predefines mesoscale pattern) + invasion OR ongoing *PHENOTYPIC* Mutations (parameters of the model)

Who persists/invades; outcompete/outevolve;

multilevel evolution: replicators and "above"

Evolution of complexity as conflict resolution

HOWEVER.....

what whe did "wrong" so far

"Defining property of biotic systems":

Very high dimensional genotype space

#### complex genotype - phenotype mapping

Therefore use of phenotypic mutations in few dimensions not appropriate

no RNA in RNA world

RNA-like Replicators dimensionless points without pysical/chemical properties

#### **Constructive Darwinian Evolution**

Darwinian evolution as efficient design (optimzation) tool

Genetic Algorithms (Holland), evolutionary computation

- population of (coded) structures/solutions/'cases'
- mutational operators
- fitness criterion
- reproduce/decay according to fitness

e.g.

computer network design/ job scheduling robotic control / body design protein design

in vitro evolution Ribozymes (RNA world)

#### in vitro RNA evolution - ligase search space ca $4^{120}$ - pop size $10^{10}$

WHY do we find it??



# Landscape important in hill climbing (and evolution of finite population(?))



Coding structure!

# RNA secondary structure as paradigm for 'natural' coding structure genotype-phenotype (GP) map

- computable 'natural' genotype-phenotype map
- RNA world
- in vitro evolution efficient

# assume fitness depends on distance to predefined target secondary structure

Early: Fontana, Schuster, Hoffacke, r Ancel, Flamm etc. (Vienna) Huynen, van Nimwegen, Takeuchi, Hogeweg (Utrecht) later/now: many others, e.g. Manrubia, Louis,....

#### **RNA Structure (tRNA-phe yeast)**



#### Computation of RNA genotype-phenotype mapping

(Zuker & Stiegler '81!)



- Base pairs
  - $\begin{cases} e(C,G) = -3\\ e(A,U) = -2\\ e(G,U) = -1\\ e(others) = 0 \end{cases}$



Otherwise,

$$E_{i,j} = \min \begin{cases} E_{i+1,j}, E_{i,j-1} & (1) \\ e(i,j) + E_{i+1,j-1} & (2) \\ \min_{i'=i+1}^{j-1} (E_{i,i'} + E_{i'+1,j}) & (3) \end{cases}$$

#### RNA secondary structure as paradigm for genotype-phenotype mapping computable??



# Min. Energy folding vs Conserved folding

A. globiformis, Anabaena sp., A. tumefaciens, B. japonicum, E. coli B. subtilis, T. thermoph, Pir. marina, Rb. sphaero

Hofacker et al 2002 Secondary Structure Prediction for Aligned RNA Sequences

### **RNA-landscape:**

## multi-one genotype-phenotype mapping

# Almost all sequences fold in 'typical shape'

but

Only small fraction of shapes is typical Example: GC strings length 30:

1.07 \* 10<sup>9</sup> sequences
218830 shapes
22718 "typical"
93.4% seqs in typical shape

Grüner, W. and Giegerich, R. and Strothmann, D. and Reidys, C. and Weber, J. and Hofacker, I.L. and Stadler, P.F. and Schuster, P. 1996

Nevertheless...

GCAU seqs. length 70: 999919 different structures in 1 million seqs



**RNA** Landscape: average phenotypic change by mutations

# (local) RNA Landscape relation distance in genotype and distance in phenotype

- single mutation: often NO change ca 30% for length 70; saturates at 20% for longer seq.
- single mutation: sometimes NO similarity (max. distance)
- distance distribution of phenotypes independent of genotype distance for moderate to large genotype distances (*small correlation length*)

# RUGGED

True for different measures of phenotypic distance Hamming distance on string representation # bond changes

# Folding of Eukaryotic mRNA: major change by point mutation (5' vs 3' end)



5

## 2 different functional ribozymes 1 point mutation NO similarity in secondary structure



shape of landscape important because of finite (localised) populaiton

- Rugged small correlation length
- identical structures overrepresented 'closeby'
- single mutation can lead to complete change of structure



-> Stuck at local optima?...NO.....

# Evolutionary dynamics of random RNA to prespecified target secondary structure





## Rugged fitness landscape

## Evolution "stuck on local optima??"









# DETOURS!

# Percolation of sequence space by neutral networks (Schuster)



Figure 10. Percolation of sequence space by neutral networks. A neutral path connects sequences of Hamming distance h = 1 (single base exchange) or h = 2 (base pair exchange) that fold into identical minimum free energy structures. The sketch shows a neutral path of length h = 9. The path ends because no identical structure was found with h = 10 and h = 11 from the reference.

# Neutral Paths (Schuster and Fontana, 1994) typical shapes percolate through shape space



Fig. 4: Neutral paths. A neutral path is defined by a series of nearest neighbour sequences that fold into identical structures.

Error/ Information threshold (as defined):

$$Q > \sigma^{-1}$$
$$L < ln(\sigma)/(1-q)$$

-->L<=0 if mutant has same fitness (phenotype) == Genotypic information threshold

> cf Phenotypic information threshold  $L < ln(\sigma)/((1-q)(1-\lambda))$

> > Takeuchi and H. 2005

Above the (genotypic) information threshold (?) (Adaptive vs) Neutral Evolution (neutral drift) (cf Kimura, theory of neutral molecular evolution)

In FLAT landscape: Diffusion through genotype space (Kimura):

$$D = 5ApL/(3 + 4pN)$$

A replication rate, p mutation rate, L length, N pop. size

On neutral network  $D' = \lambda D$ 

#### evolution over neutral network is diffusion-like process



measured diffusion in RNA landscape (in target structure)

# Higgs and Derrida: for finite populations "speciation" in flat landscape



Fig. 1. Distribution of the elements of the matrix  $T^{\alpha\beta}$  in the OPM for a population of M = 1000 individuals. The distribution is shown at six times for the same population. There is a period of 50 generations between each successive pair of curves; therefore the peaks move a distance 50 to the right each time. Peaks fluctuate in size and eventually disappear.



Fig. 2. Schematic representation of the genealogical to OPM showing ultrametric property of the branching t  $T^{\alpha\gamma}$ ,  $T^{\beta\gamma}$ . Cutting the tree at an arbitrary point in the pathe population into families.

## "punctuated evolution" ("epochal evolution")



### **Punctuated evolutionary dynamics**

(vs "new synthesis" vs Gould)

- external environmantel change???
- "waiting for unlikely mutation" stuck on local optimum
- ecological quillibrium stable spatial patterns
- phenotypic punctuated equillibria stasis while on neutral path

#### **Evolutionary dynamics: population structure**

400 Evolution: Huynen et al.

Proc. Natl. Acad. Sci. USA 93 (1996)



FIG. 3. Evolutionary optimization. A flow reactor with capacity N = 1000 is initialized with that many copies of a random sequence of length  $\nu = 76$ . The mutation rate is p = 0.001 and the target secondary structure is the tRNA<sup>Phe</sup> cloverleaf, the replication rate function is  $A(d) = 1.06^{146-d}$ , where *d* is the tree-edit distance (9) to the target structure. The population average of the distance to the target is plotted against time (solid line) for a specific interval of the entire run (*Inset*). Superimposed series of dots render the evolution of the population structure over time. Dots at one time epoch are a one(!)-dimensional projection (see Fig. 2 legend) of the population of sequences present in >10 copies at that time. Collecting and random target structures we tried for both linear and exponential fitness functions A(d).

#### **Population Structure: landscape sampling**



FIG. 2. Population structure in sequence space. The support of a population in sequence space is the set of sequences present in at least one copy. The population support can be pictured in two dimensions using some theorems from distance geometry (27). We compute the metric matrix *M* with entries  $m_{ij} = (d_{ij}^2 + d_{ij}^2 - d_{ij}^2)/2$ , where  $d_{ij}$  is the Hamming distance between sequences *i* and *j* and 0 is the center of mass of the support. Sequences are expressed in principal axes coordinates by diagonalizing *M*. Only the components corresponding to the largest two eigenvalues are kept, yielding a projection onto the plane that captures most of the variation. Dots represent a static snapshot of N = 2000 individuals after 135 time units replicating with p = 0.002. Among the 2000 individuals, 631 are different and among them 301 fold into different structures. To help correct for the distortions of the projection, the dots are connected by the edges of the minimum spanning tree. Edges connect closest points. Red (blue), Hamming distance less (more) than 6; dot size large (small), more (less) than four copies in the population; yellow (green), sequences that do (do not) fold into the tRNA target structure.

#### Novelty "seen" along the neutral path (Huynen 1998)



Innovations'

Shadow of similar structures along neutral paths

Zuckerhandl "Neutral + adaptationist evolution reconciled" (Kimura memorial lecture)

# RNA Genotype - Phenotype mapping Ideal for evolution

(Schuster and Fontana, 1994)



Sequence Space

Shape Space

Fig. 5: A sketch of the mapping from sequences into RNA secondary structures as derived here. Any random sequence is surrounded by a ball in sequence space which contains sequences folding into (almost) all common structures. The radius of this ball is much smaller than the dimension of sequence space.

# Shape of RNA fitness landscape percolating and intertwining Neutral Networks:



#### closer look.... complete neutral network from AUACGAAACGUA (1094 genotypes) + connections of 2 novel phenotypes to the network



note clustering of grey neutral network and of portals to red and blue Non-Poissonian Bursts in the Arrival of Phenotypic Variation Can Strongly Affect the Dynamics of Adaptation Nora Martin . Art louis 2024

### "arrival" of novel phenotypes non random distrubuted"



(measured under strong slection for grey phenotype

### Also for longer sequences (L=30)



under weak selection, bursts increase chance of invasion of novel phenotypes

### **MOREOVER:** phenotype – > function mapping



Alternative ligases (Ekland et al 1995)

'tyranny' of small motifs... or complex structures?

## 'drift' on neutral network not 'neutral': (1) Longterm RNA evolution: fitness of mutants



Evolutionary target of 2 hairpin loops, of maximum length (fitness =  $(L1 \times L2)$ . Huynen & Hogeweg 1993)

## (2) Evolution towards high lambda



# redundant genotype-phenotype mapping: choice of coding

- Evolution towards 'flatter parts'
  - == Mutational robustnes
  - == high connectivity of neutral network
  - == MAX EIGENVECTOR OF CONNECTION MATRIX
  - == D= Max eigen value
  - (van Nimwegen 2000)

compare blind ant (moves with prob. rel neutral NB)

$$--$$
 > same freq in each node)

myopic ant (moves with fixed probability)  $-- > D = \hat{d} + Var(d)/\hat{d}$  Evolution towards mutational robustness == largest eigenvalue of connection matrix van Nimwegen et al PNAS 1999



#### walk along neutral path not neutral

# walk along neutral path not neutral..... how neutral is neutral



# walk along neutral path not neutral..... how neutral is neutral



neutral if above the informatioon threshold!

# example of intra-molecular evolved landscape negative epistasis



Hsp90,582-590 Effect on growth-rate of single point mutations from wild-type and from 7 (almost) neutral mutations

A systematic survey of an intragenic epistatic landscape Claudia Bank et al MBE 2014