

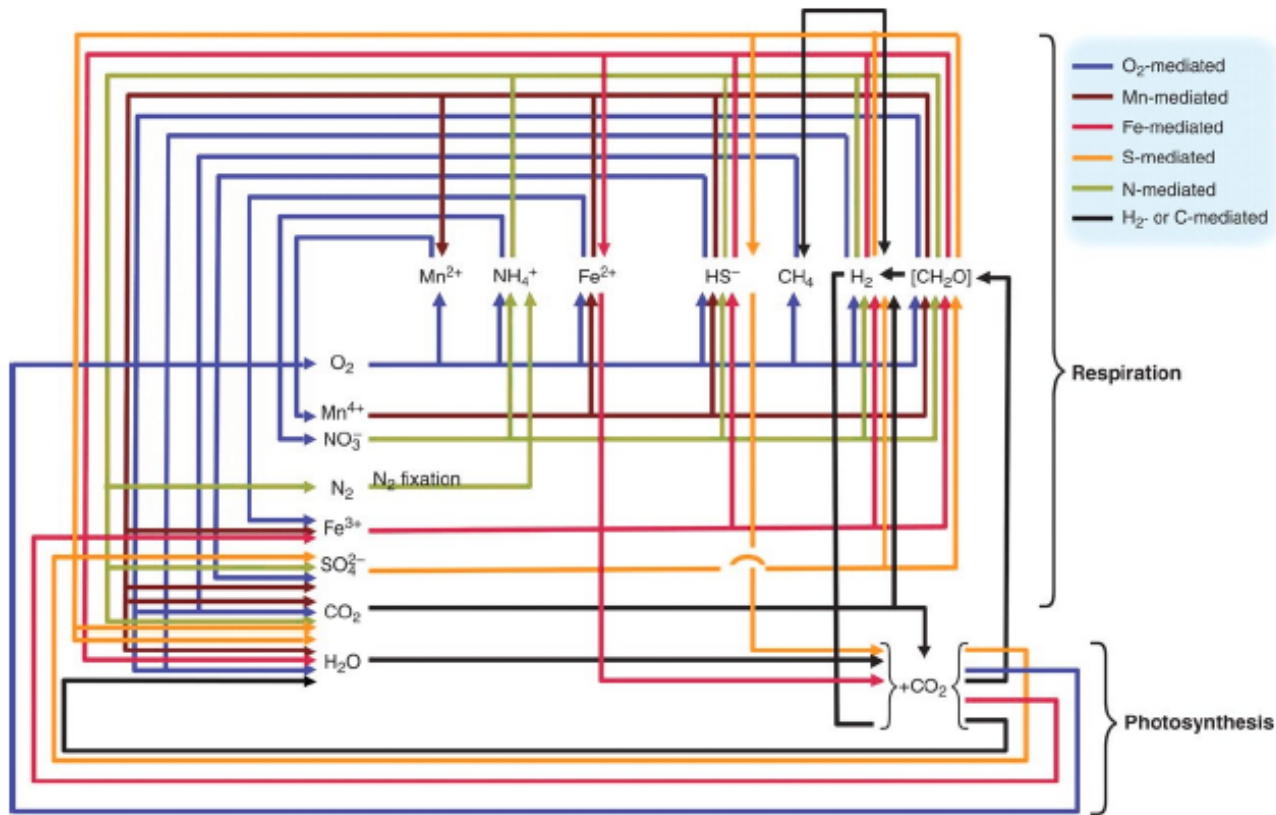
13b

Metabolism: flux balance analysis

Course computational Biology 2024; Paulien Hogeweg;

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(1) Life *is*..... energy/nutrient cycling



“The individual taxonomic units evolve and go extinct, yet the core machines survive surprisingly unperturbed.”

PG Falkowski et al, Science 2008

Metabolic networks: Exploiting constraints

metabolic network are evolved

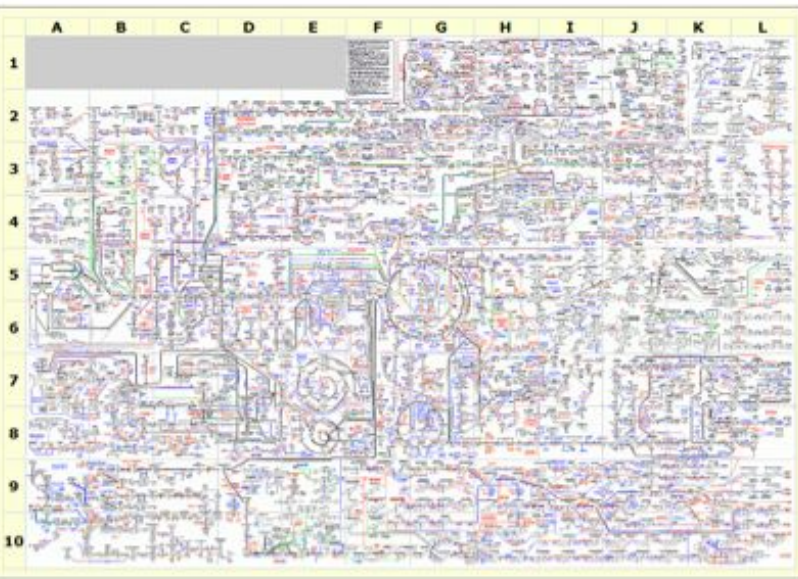
However metabolic networks many physical/chemical constraints

stoichiometry, energetic constraints

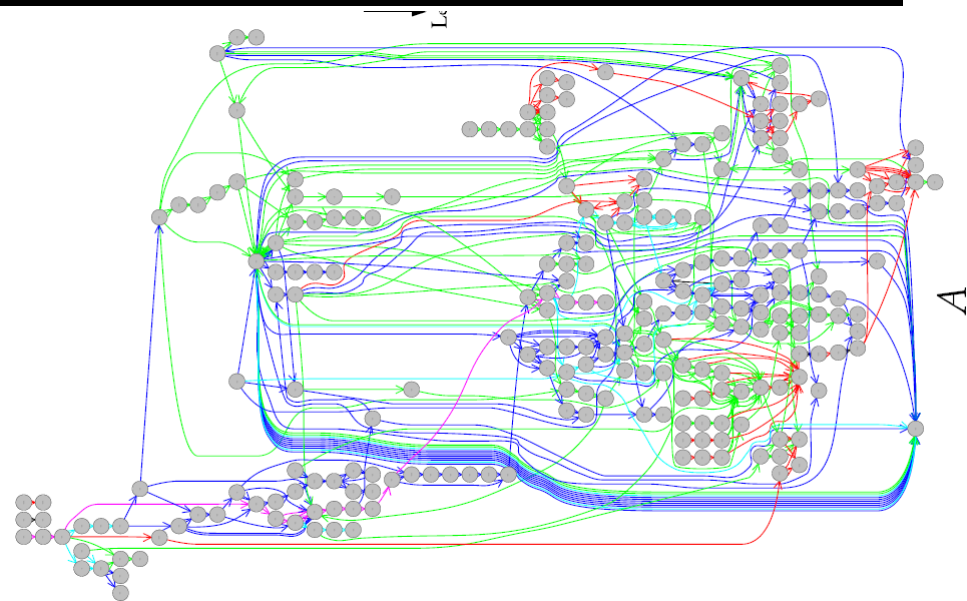
can/should be exploited

allows for model upscaling to complexity of present day organisms

stoichiometric constraint, + equilibrium assumption
allows calculation of (optimal) flux through large
metabolic networks

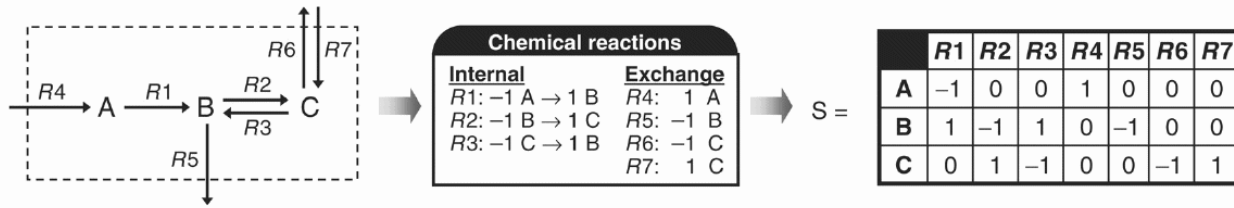


KEGG database



metabolic network yeast

I. Reaction network formalism



II. FBA formulation

Dynamic mass balance

$$\frac{dC}{dt} = Sv$$

C : Concentration
t : Time
S : Stoichiometric matrix
v : Flux vector

Steady-state assumption

$$Sv = 0$$

LP formulation

Objective: $\max Z = v_5$

Constraints:

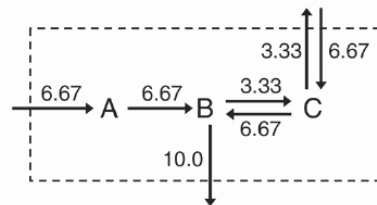
$$\begin{array}{c}
 A \\
 B \\
 C
 \end{array}
 \begin{bmatrix}
 R1 & R2 & R3 & R4 & R5 & R6 & R7 \\
 -1 & 0 & 0 & 1 & 0 & 0 & 0 \\
 1 & -1 & 1 & 0 & -1 & 0 & 0 \\
 0 & 1 & -1 & 0 & 0 & -1 & 1
 \end{bmatrix}
 \begin{bmatrix}
 v_1 \\
 \vdots \\
 v_7
 \end{bmatrix}
 = 0$$

$0 \leq v_1, \dots, v_7 \leq 10$

III. Hypothetical flux distribution at steady-state

$$Z = 10$$

$$v = [6.67 \ 3.33 \ 6.67 \ 6.67 \ 10.0 \ 3.33 \ 6.67]^T$$



Flux balance analysis FBA

assume 'automatic' regulation such that flux in equilibrium and maximal growth

- FBA solution non-unique:
use secondary optimization, eq minimal total flux
- stoichiometric matrix ($n_A - > m_B$)
- reactions coupled to enzymes
- set maximum flux (when enzymes are present) (OR, AND reactions))
- however actual flux not proportional to amount of enzymes

examples of flux balance analysis

- How do fluxes (growth) change with change of environment (=input-flux)
- How do fluxes (growth) change with knock-outs?
- How do fluxes (growth) change after endosymbiosis? (cf Pal Papp Nature 2006)
- reconstruction of Ecosystem wide metabolome (cf Bas Dutilh)
-
- **How do genomes reduce after whole genome duplication?**
(cf van Hoek and Hogeweg 2009)

**automatic reconstruction of metabolic networks from
annotated genomes of Henry CS1, DeJongh M, Best
AA, Frybarger PM, Linsay B, Stevens RL. 2010**

High-Throughput Genome-scale Metabolic Reconstruction Pipeline

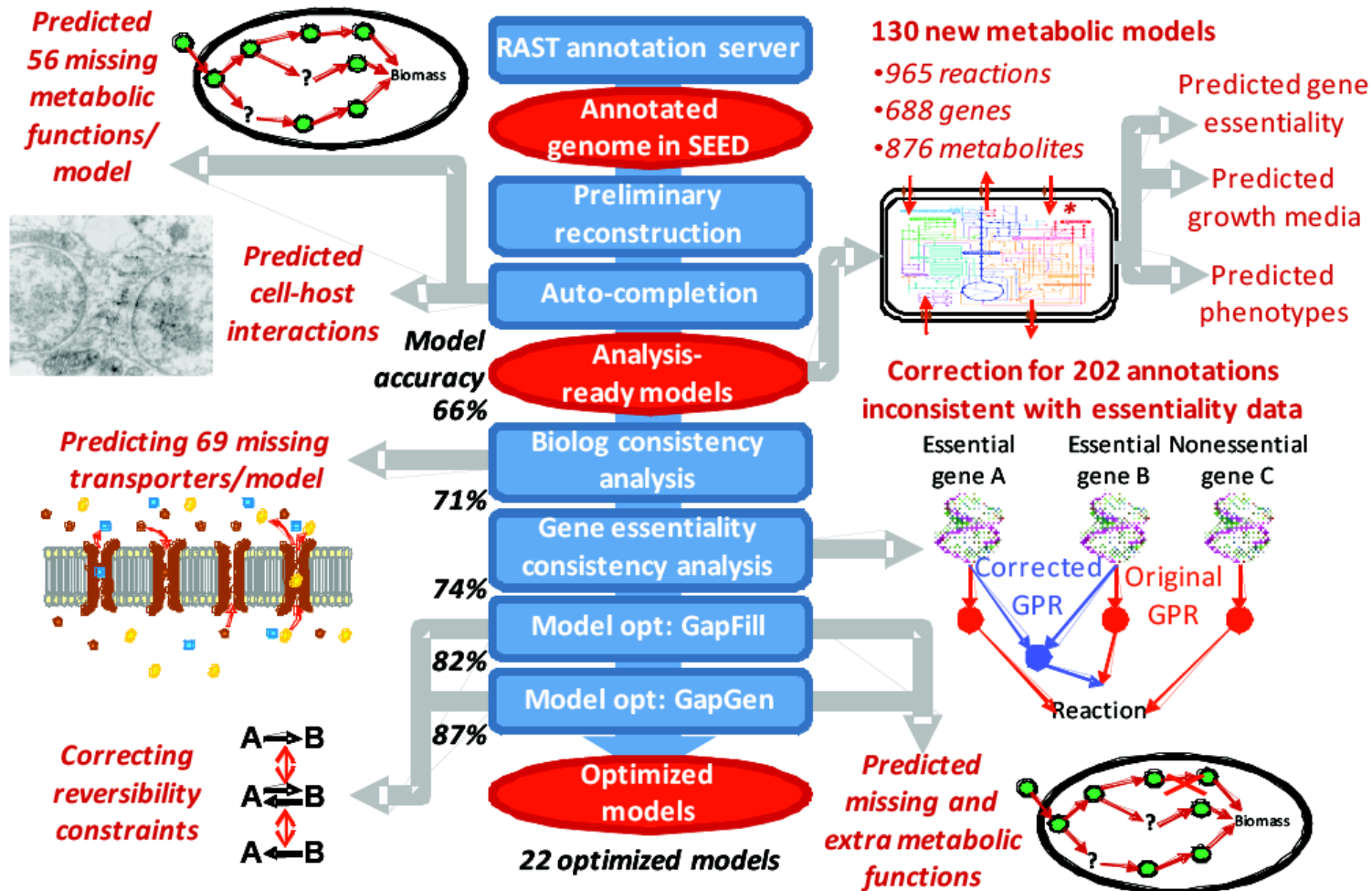
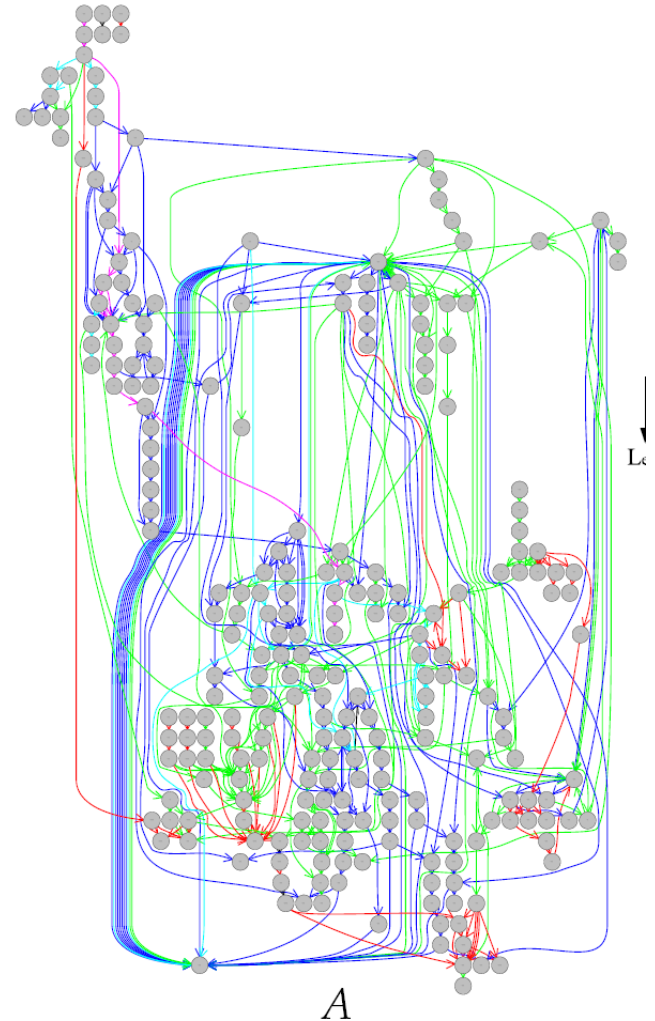


Figure S1: Overview of results and discoveries arising from the Model SEED pipeline.

Using FBA to reconstruct evolution of metabolic network of yeast after WGD

to cope with
genome-size networks:
exploit constraints
and use shortcut:
optimal equilibrium flux



Yeast metab. network

evolution of metabolic flux after WGD

FBA assumptions

- WGD – > volume increase (decrease surface/volume ratio)
volume = depends on genome size
- flux of metabolic reaction depends on gene expression,
dosis effect: gene copy number
- max flux through each reaction preset to maximum needed
for optimal growth in sampled set of realizations of 10 en-
vironment types
- enzymes have multiple functions

- reactions need multiple enzymes
take into account OR, AND (AND/OR) relations
- flux transport reactions: depends on gene expression AND
surface/volume ratio
- after gene deletion maxflux reduced accordingly

WGD, cell size and fluxes

cell size scales with amount of DNA

Cavallier Smith (e.g. 2005)

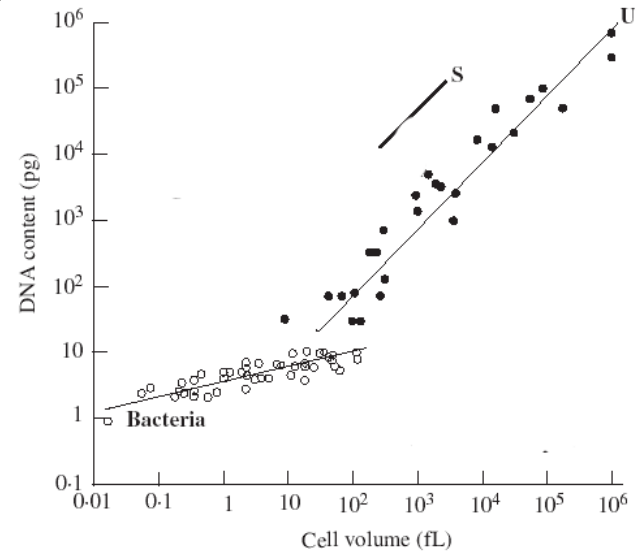
In Yeast diploide cells are:

$V=1.89 * \text{haploid cells}$

surface: $1.56 * \text{haploid cells}$

$V = N \cdot 9; A = V \cdot 7$

where N number of genes



MaxFlux change as function of area change (α), volume change

(β) and gene dosage change γ)

external flux

$$F_{\max}(i) = F_{\max,0}(i) \frac{\alpha \gamma(i)}{\beta} \frac{1+x(i)}{\gamma(i)x(i) + \alpha}$$

internal flux

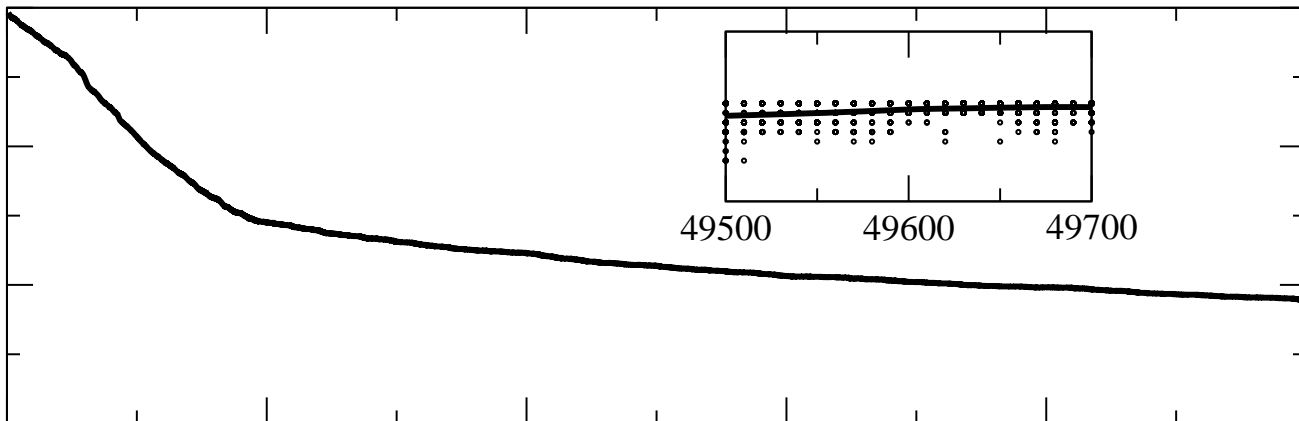
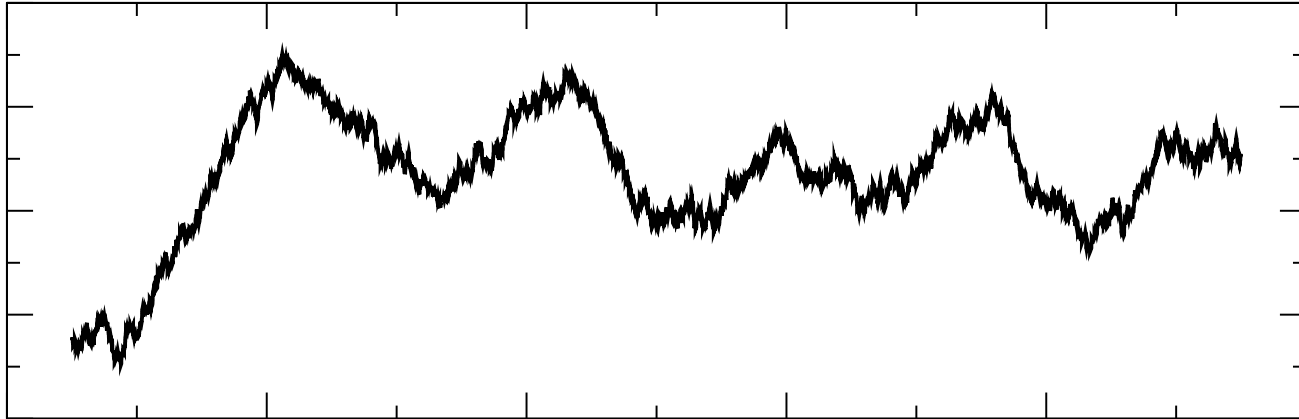
$$F_{\max}(i) = F_{\max,0}(i) \frac{\gamma(i)}{\beta}$$

evolution of metabolic flux after WGD evolutionary protocol

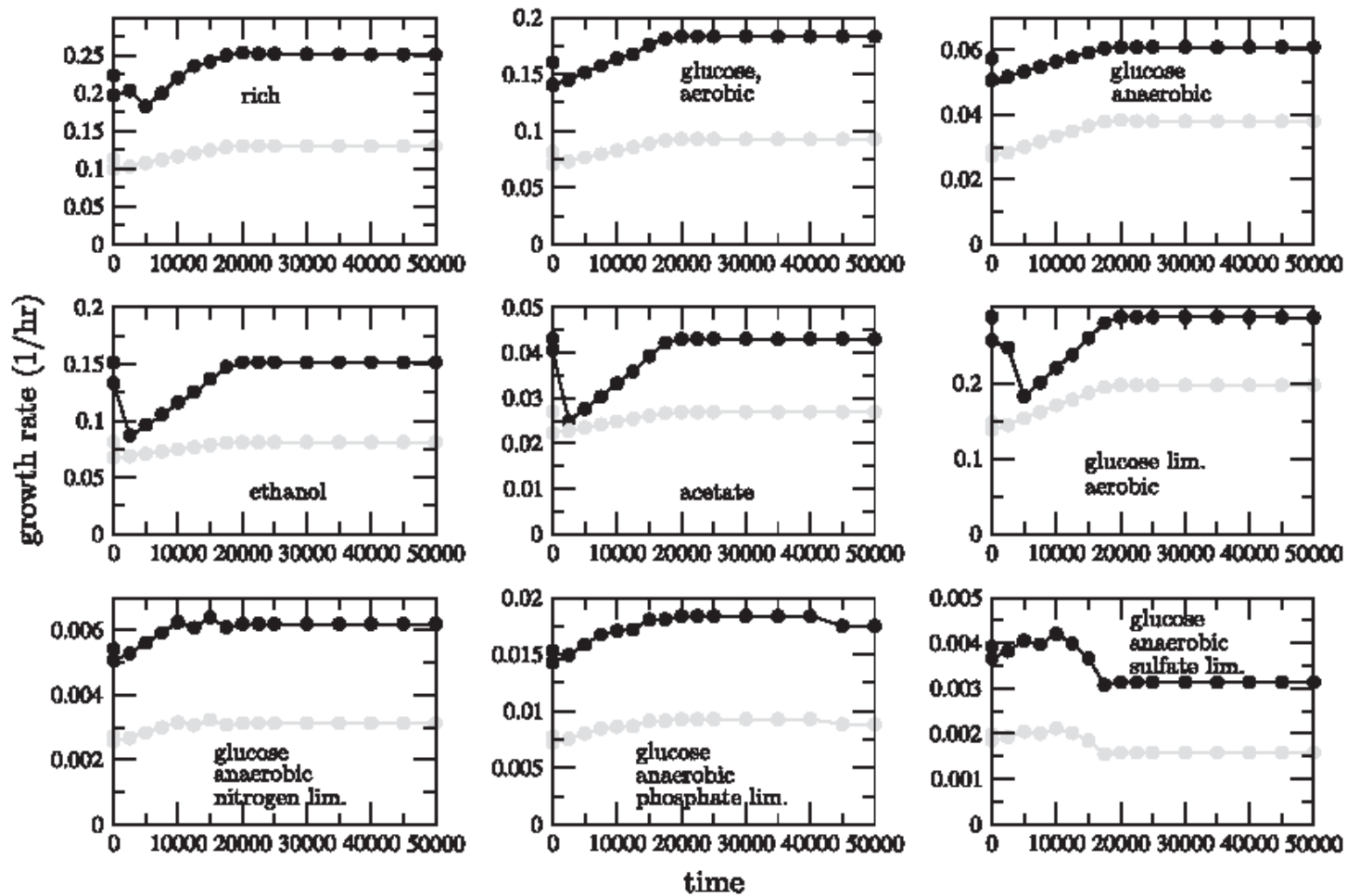
- 9 types of environments (available nutrients).
realized in different concentrations
- per generation 1 environment seen
- pop size 100: flux dependent replication
death: no growth + random
- after wgd: only deletions
or duplication + deletion (max 2 copies)
- no fitness advantage for genome shrinkage smaller than
initial volume

evolution of metabolic flux after WGD

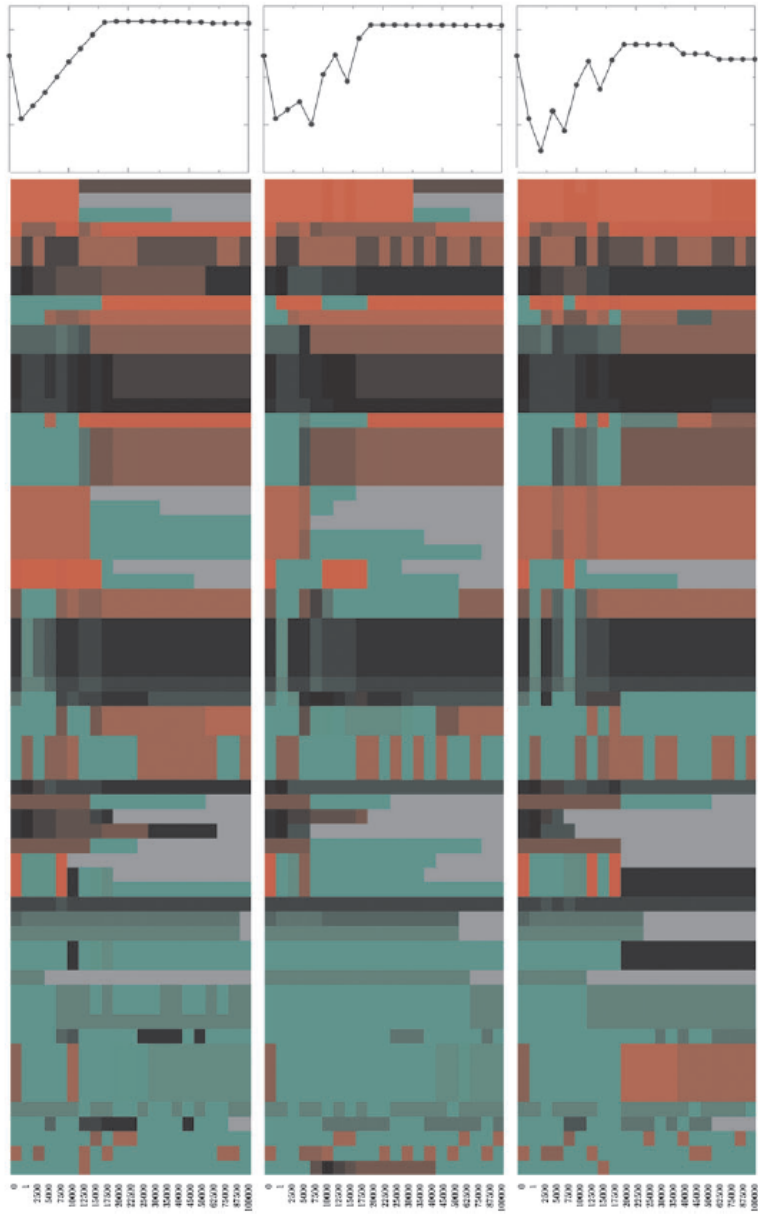
evolutionary dynamics: growth rate and genome reduction



**evolution of metabolic flux after WGD
flux in the various environments
(max and mean concentration)
initial decrease – how/when does it happen in
evolution**



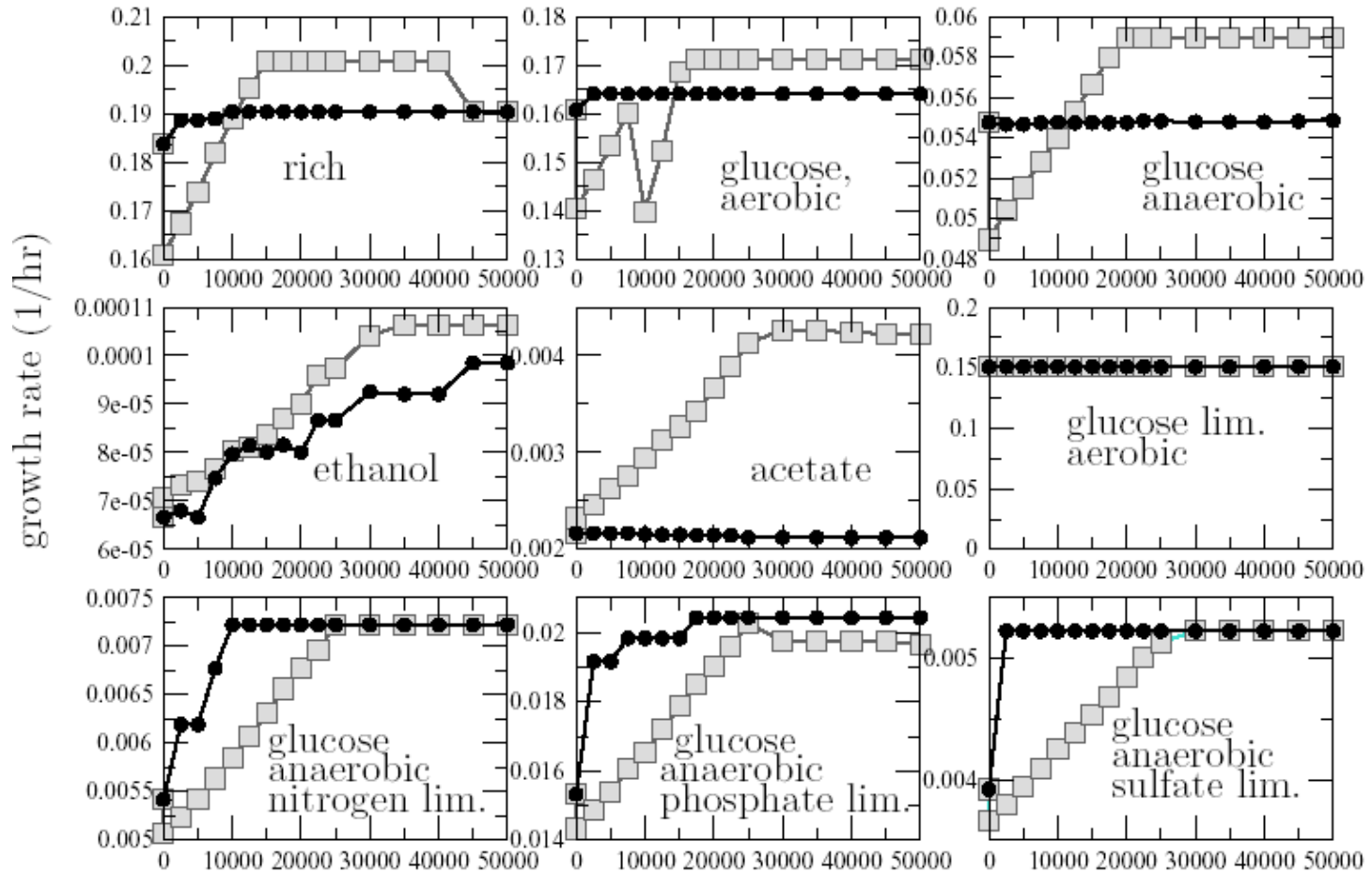
**Genome shrinkage after whole genome duplication
dynamics of use of pathways in anaerobic glucose
environment (env 3)**



0.06
0.05

- 618 Glycerolipid_Metabolism
- 687 Transport_Extracellular
- 621 Glycerolipid_Metabolism
- 1101 Transport_Mitochondrial
- **183 Alanine_and_Aspartate_Metabolism
- **187 Transport_Mitochondrial
- 336 Transport_Mitochondrial
- 612 Oxidative_Phosphorylation
- 686 Oxidative_Phosphorylation
- 1185 Threonine_and_Lysine_Metabolism
- 278 Cysteine_Metabolism
- 1139 Cysteine_Metabolism
- 96 Nucleotide_Salvage_Pathway
- 109 Methionine_Metabolism
- 849 Methionine_Metabolism
- 296 Phospholipid_Biosynthesis
- *78 Transport_Extracellular
- 278 Methionine_Metabolism
- 1148 Methionine_Metabolism
- 1149 Methionine_Metabolism
- 1186 Threonine_and_Lysine_Metabolism
- 180 Alanine_and_Aspartate_Metabolism
- 763 Glycine_and_Serine_Metabolism
- 169 Alanine_and_Aspartate_Metabolism
- 1198 Threonine_and_Lysine_Metabolism
- 764 Glycine_and_Serine_Metabolism
- 620 Glycerolipid_Metabolism
- 619 Glycerolipid_Metabolism
- 14 Transport_Mitochondrial
- 801 Valine_Leucine_and_Isoleucine_Metabolism
- 657 Glycine_and_Serine_Metabolism
- 693 Transport_Mitochondrial
- 876 Transport_Mitochondrial
- 1143 Transport_Mitochondrial
- 763 Methionine_Metabolism
- 1235 Transport_Extracellular
- 200 Transport_Mitochondrial
- 1022 Transport_Mitochondrial
- **12 Transport_Mitochondrial
- **1225 Tyrosine_Tryptophan_and_Phenylalanine_Metabolism
- **1229 Transport_Mitochondrial
- 1169 Citric_Acid_Cycle
- 112 Methionine_Metabolism
- 59 Pyruvate_Metabolism
- 677 Glutamate_Metabolism
- 763 Methionine_Metabolism
- 259 Transport_Mitochondrial
- 269 Alanine_and_Aspartate_Metabolism
- 264 Alanine_and_Aspartate_Metabolism
- 574 Fatty_Acid_Biosynthesis
- 1120 Nucleotide_Salvage_Pathway
- 1116 Nucleotide_Salvage_Pathway
- 73 Transport_Mitochondrial
- 262 Transport_Mitochondrial
- 301 Purine_and_Pyrimidine_Biosynthesis
- 909 Nucleotide_Salvage_Pathway
- 1124 Nucleotide_Salvage_Pathway
- 288 Nucleotide_Salvage_Pathway
- 1168 Oxidative_Phosphorylation
- 76 Pyruvate_Metabolism
- 79 Transport_Mitochondrial
- 89 Nucleotide_Salvage_Pathway
- 1038 Oxidative_Phosphorylation
- 658 Purine_and_Pyrimidine_Biosynthesis
- 602 Oxidative_Phosphorylation
- 1223 Tyrosine_Tryptophan_and_Phenylalanine_Metabolism
- 1224 Tyrosine_Tryptophan_and_Phenylalanine_Metabolism
- 782 Transport_Extracellular

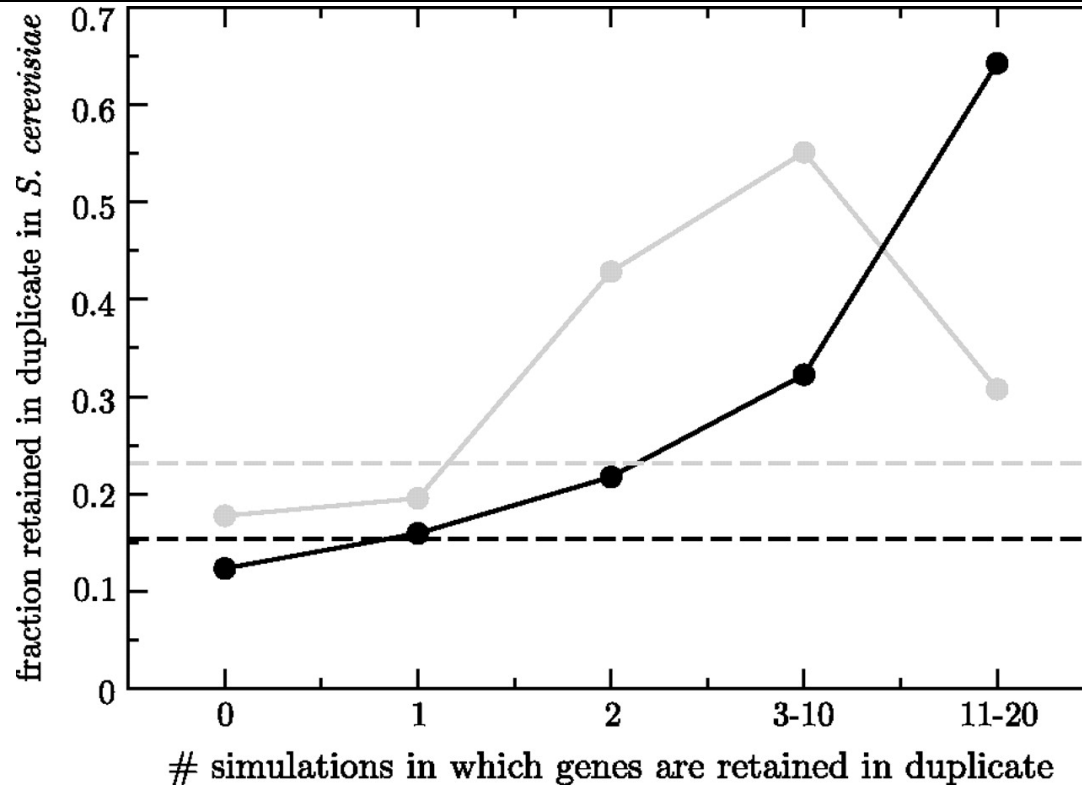
Only in “new” environment - nodirect disadvantage of WGD



BUT single INDELS initially better Exept in ethanol env

WGD mostly better end result than single INDELS

WGD: Simulated evolution and/vs yeast duplication of yeast vs duplication of ancestor of yeast (+hgt)



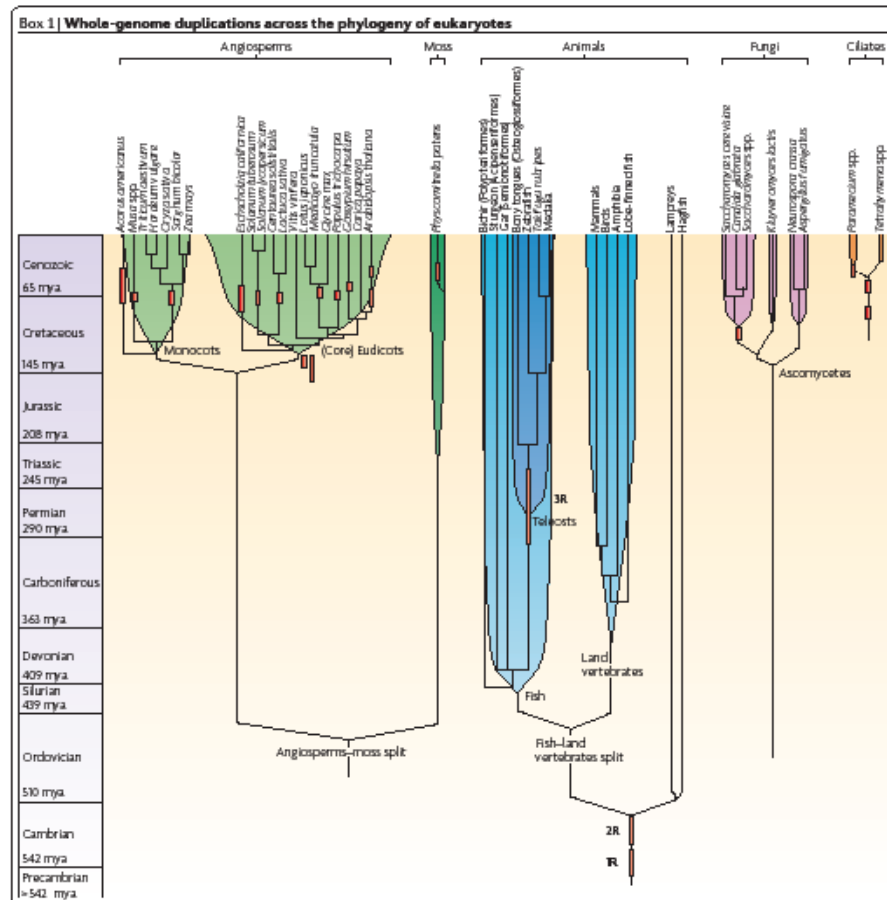
Preferential retained genes: Glycolysis pathway and Transporters

Evolution predictable!

conclusions
supervised vs non-supervised modeling of
WGD in Yeast

“Supervised” Conan & Wolfe (2007)	“Non Supervised” van Hoek & H. (2009)
find genes preferentially retained <i>glycolysis pathway</i> Model glycolysis pathway assuming dosis effect of duplicated genes demonstrate WGD can lead to increased glycolic flux	take known interactions metab. net + DNA-volume relation model evolution find preferentially retained genes <i>glycolysis & transport</i> WGD mostly disadvantageous initially except in “new” environments seldom better than single INDELS evolutionary outcome “deterministic”
WGD enabled to exploit high glucose resource during emergence of angiosperms	WGD enabled to exploit high glucose resource during emergence of angiosperms
observed outcome of WGD	expected outcome of WGD

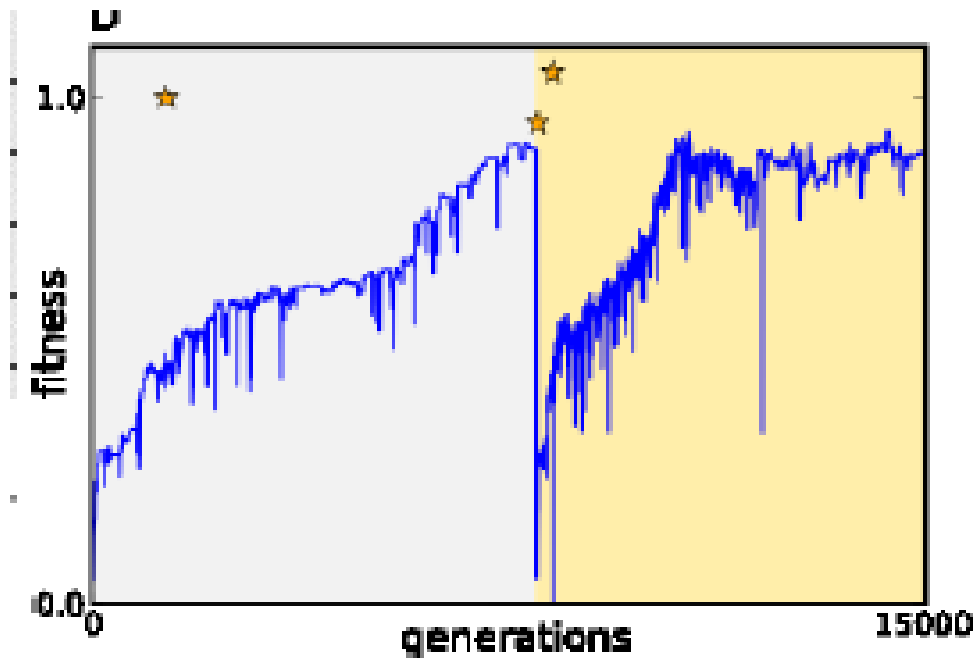
WGD observed in phylogeny at times of environmental shifts



van der Peer et al 2009, Nature genetic reviews

WGD observed in virtual cell model at times of environmental shifts

WGD ongoing mutation,
but only fixed in population EARLY in evolution
OR after SOME (severe?) environmental changes



and WGD leads to high fitness much later

Cuypers & Hogeweg 2014

Neutral Paths, Causal Drift, Robust Signaling, and Complex Disease

Andreas Wagner 2015 PLONE

Explicit model of Insuline signaling pathway

Random sampling of 15 kinetic parameters $10^{-3} - 10^3$ and evolving populations by mutating these parameters

Generate many “healthy” and “sick” individuals (pathway instantiations)

Classifying behavior as “normal” $V = 0.076x10^{-4}$
or “deseased” $V = 0.33x10^{-4}$

(based on glucase uptake-curve in time) Determine sensitivity of parameters in different populations and during evolution.

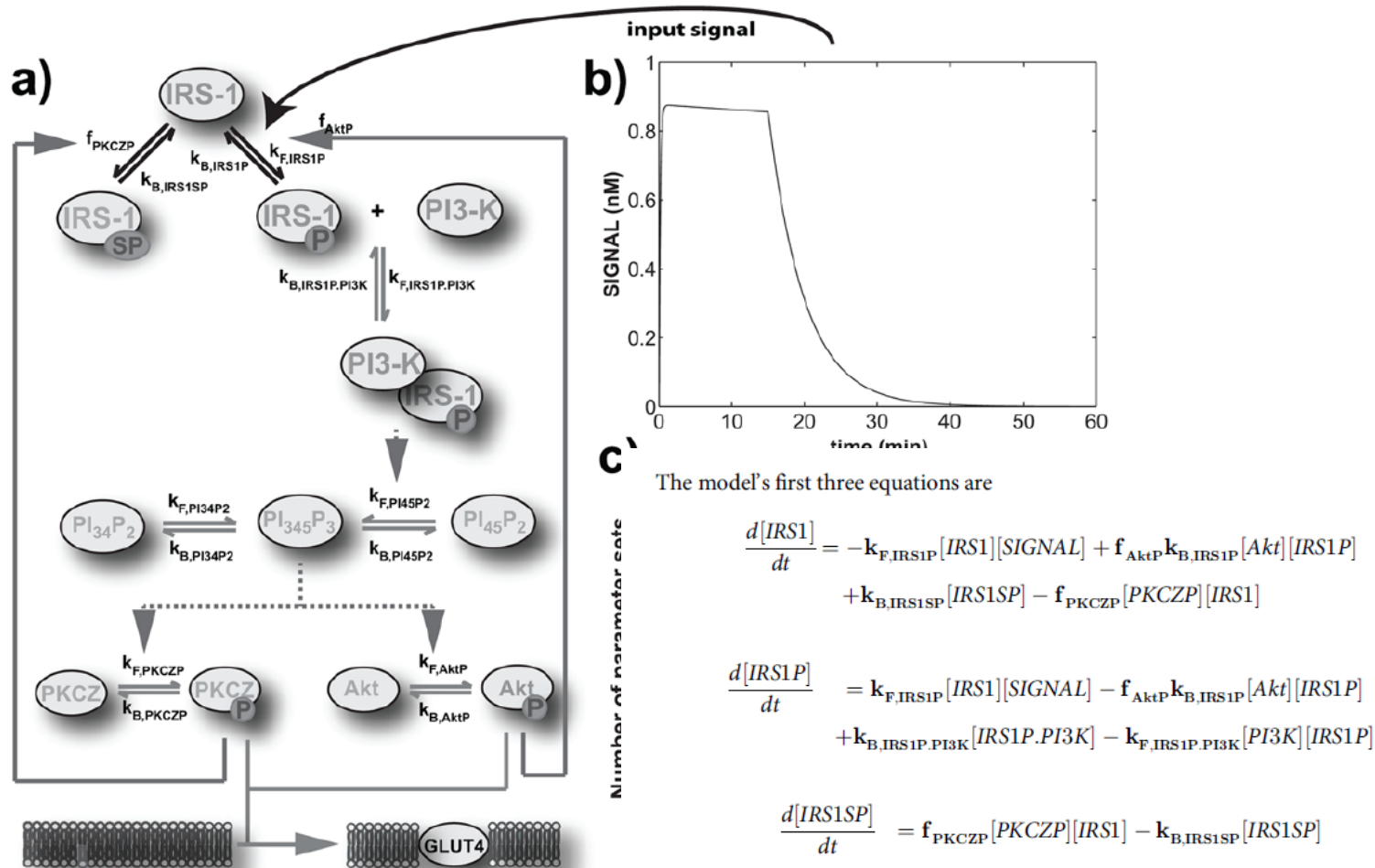


Fig 1. Insulin signaling model, input and output. a) Molecular interactions in the signaling pathway modeled here. Briefly, extracellular insulin leads to phosphorylation of the insulin receptor, which promotes the phosphorylation of *IRS1* to yield *IRS1P*. The latter molecule associates with *PI3K* in a complex that triggers production of the second messenger $PI_{345}P_3$, which activates the protein kinases *Akt* and *PKCZ*. These kinases then promote the translocation of the glucose transporter *GLUT4* to the membrane, where it helps import glucose into the cell. Mass-action parameters that determine the rates of the respective reactions are indicated by a 'k' followed by a subscript. Activated *PKCZ* and *Akt* exert feedback on the production of two different phosphorylated forms of *IRS1* (*IRS1SP* and *IRS1P*). The strength of this feedback is encapsulated by parameters f_{PKCZP} and f_{AktP} , respectively. See [Methods](#) for details. b)

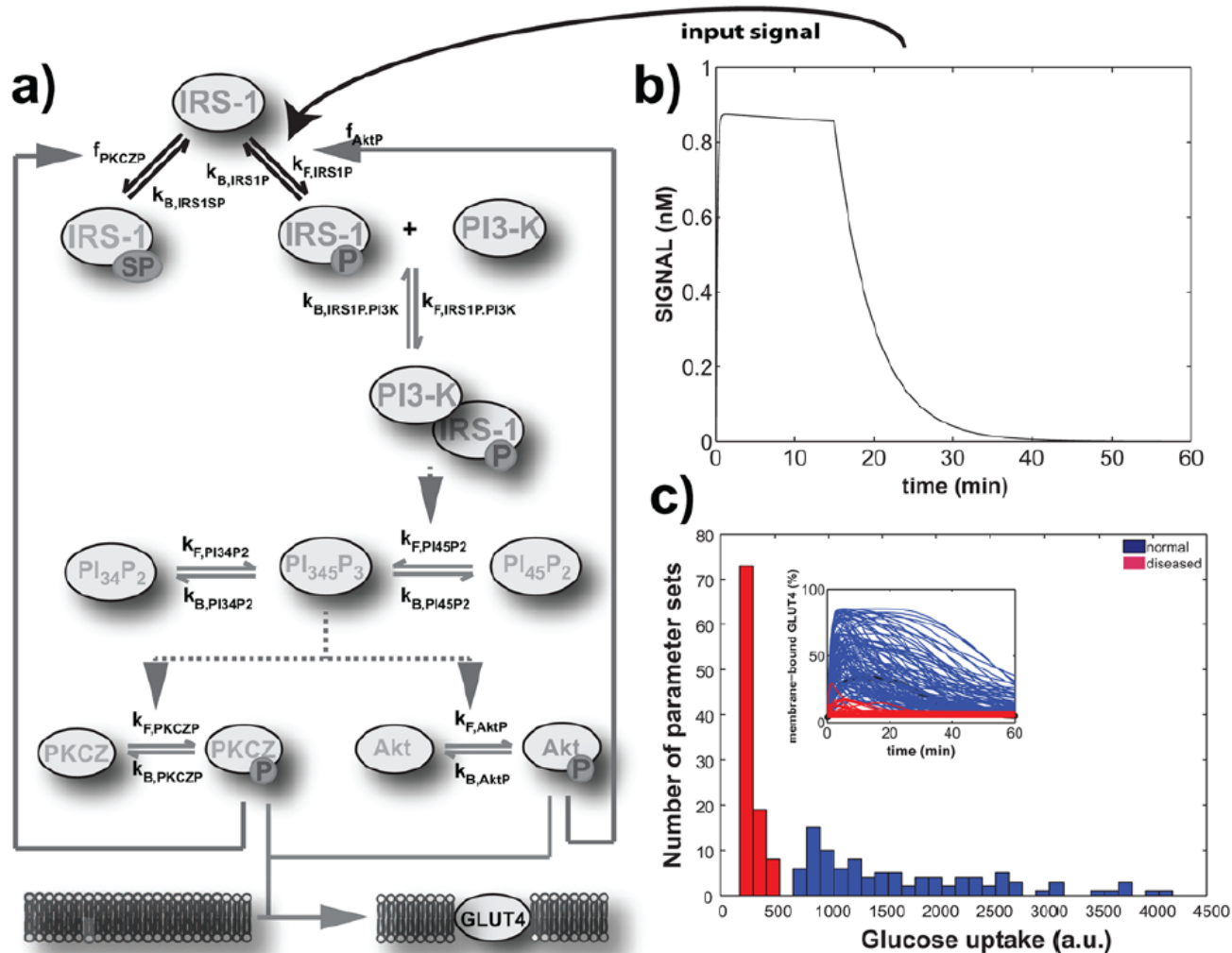
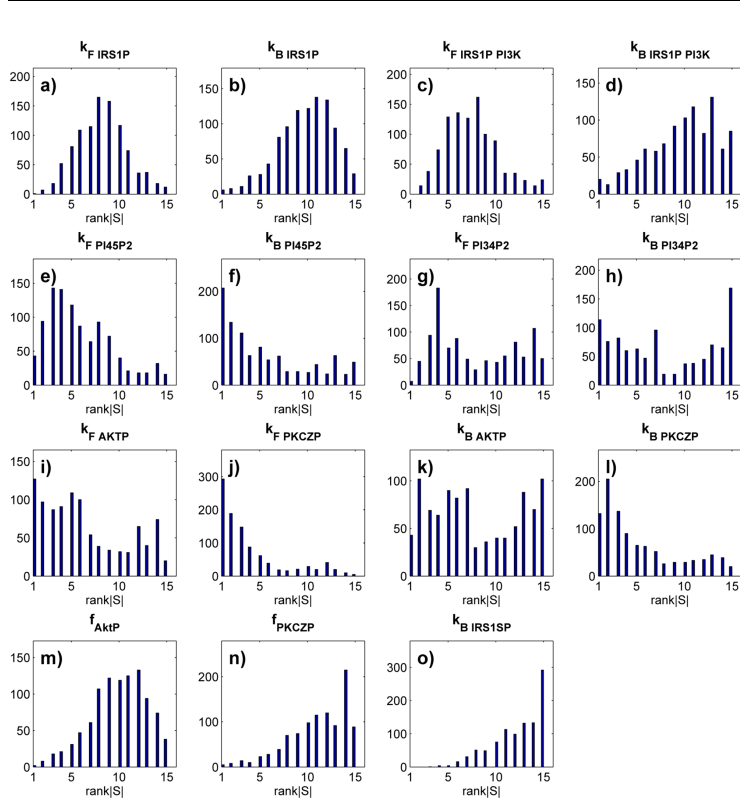
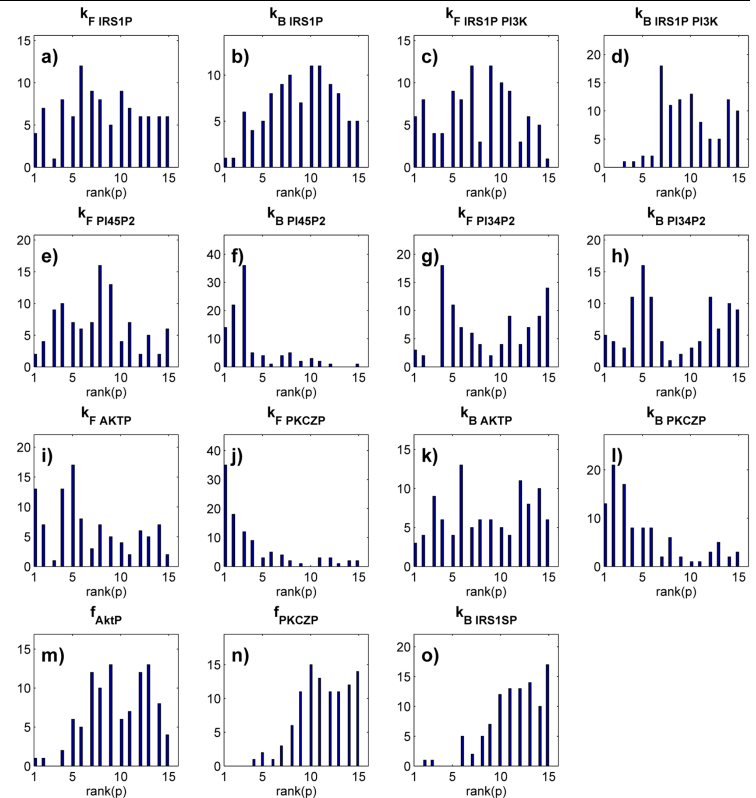


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Importance of parameter varies greatly dependment om parameter set (= genetic background)

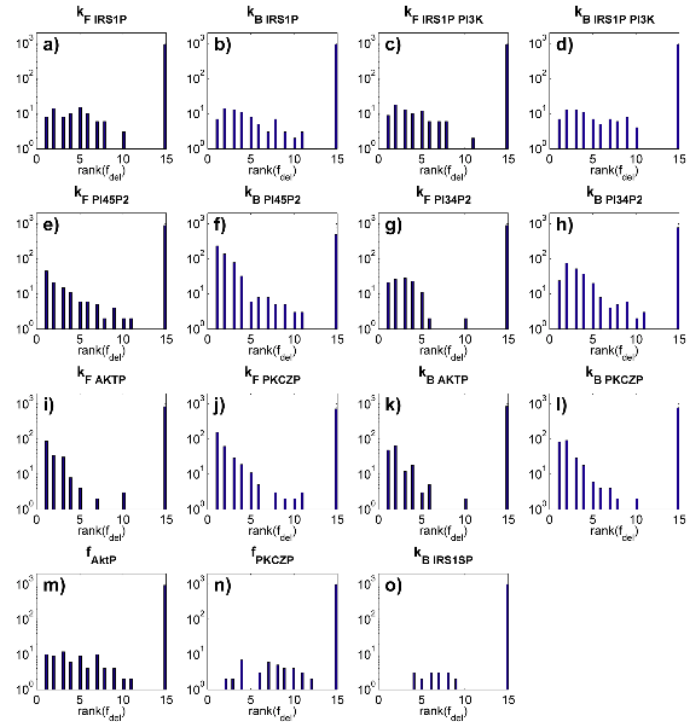
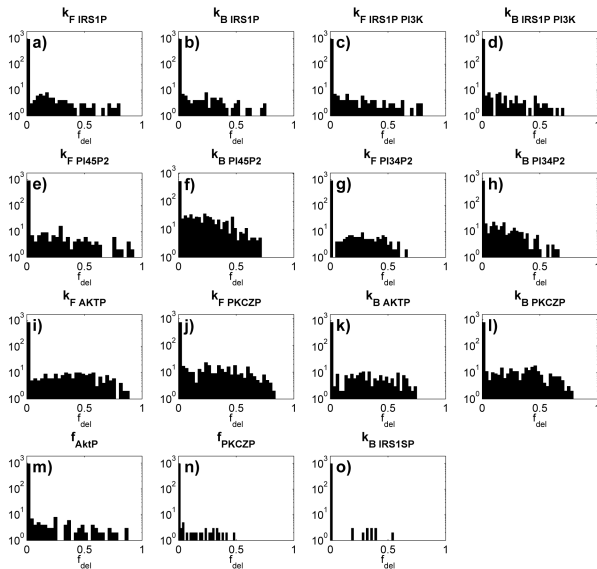


sensitivity of parameter



discrimination healthy/sick

very high neutrality of 'gene' deletions
 but very different in different parameter sets
 (instantiations).



neutrality of deletions

likelihood of deleterious effects

Rapid “Causal drift”

rapid change of sensitivity to parameter changes (mutations) due to neutral drift

“genetic background”

“cause of disease”

cf GWAS studies
50% “explained”

Mouse models

